CHM 4300L, Laboratory in Biochemistry & Molecular Biology

Summer 2018

Professor Jon D. Stewart

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Office hours Tuesday, 3:30 - 4:30 p.m.

Thursday, 9:00– 10:00 a.m. Friday, 11:00 a.m. – 12:00 p.m.

Teaching Assistants Nasser Faghih (nasseruf@chem.ufl.edu)

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Course Objectives CHM 4300L provides a hands-on opportunity to learn basic techniques

in molecular biology and biochemistry including the polymerase chain reaction, DNA cloning, plasmid isolation and characterization, protein

purification and steady-state kinetics.

Prerequisites CHM 2211, CHM 2211L and either CHM 3218 or BCH 4024.

Grading Three lab reports (100 points each) will be due over the course of the

semester [due on June 20 or 21 (depending on section), July 18 or 19 (depending on section) and August 10 (both sections)]. Guidelines for each lab report will be distributed at least one week before each due date.

Lab notebooks will also be graded at these times for accuracy and completeness (100 points total). Four short (20 minute) quizzes will be given at the beginning of pre-lab lectures at unannounced times during the semester (total 100 points). *These quizzes will cover material for that week's lab only*. The experiments in this course are arranged in a series, and the product from one week serves as the starting material for the next. If you have problems, you will be provided with intermediate

materials with no grade penalty. You will be expected to analyze critically where the problem(s) lay in your lab report, however, and this will be graded (see above). Current UF grading policies can be found at

https://catalog.ufl.edu/ugrad/current/regulations/info/grades.aspx.

Class Attendance Attendance is required at all laboratory sessions; please be on time!

Attendance at pre-lab lectures is strongly suggested. Please do not schedule school visits, interviews, etc. during lab periods. Excused absences are consistent with university policies in the undergraduate

catalog

(https://catalog.ufl.edu/ugrad/current/regulations/info/attendance.aspx)

and require appropriate documentation.

Make-Up Work Because they are unannounced, there are no make-up quizzes. Students

with an allowable excuse should see the Instructor to arrange

accommodations if they miss a quiz. Late reports and lab notebooks will not be accepted for grading.

Students are expected to provide feedback on the quality of instruction in

Course Evaluation

this course by completing online evaluations at https://evaluations.ufl.edu. Evaluations are typically open during the last two or three weeks of the semester, but students will be given specific times when they are open. Summary results of these assessments are available to students at https://evaluations.ufl.edu/results/.

Class Demeanor

Students are expected to arrive to class on time and behave in a manner that is respectful to the instructor and to fellow students. Please avoid the use of cell phones and restrict eating to outside of the classroom. Opinions held by other students should be respected in discussion, and conversations that do not contribute to the discussion should be held at minimum, if at all.

Required Textbook

Cloning, Expression & Characterization of Human Carbonic Anhydrase II. Experimental Manual. Summer 2018 Edition (available at Target Copy Center).

Laboratory Schedule

A calendar of scheduled experiments is available at the Canvas e-Learning site (http://elearning.ufl.edu).

Laboratory Attire

Department of Chemistry approved SAFETY GLASSES or GOGGLES (only Astro OTG 3001 or American Optical 91214 Goggles) must be worn any time you are in the laboratory. Protective clothing (long pants, shirt/blouse and proper shoes) is required any time you are in the laboratory and it must be worn at all times while in the laboratory. A more detailed discussion of laboratory safety is available on the course e-Learning site (http://lss.at.ufl.edu).

Academic Honesty

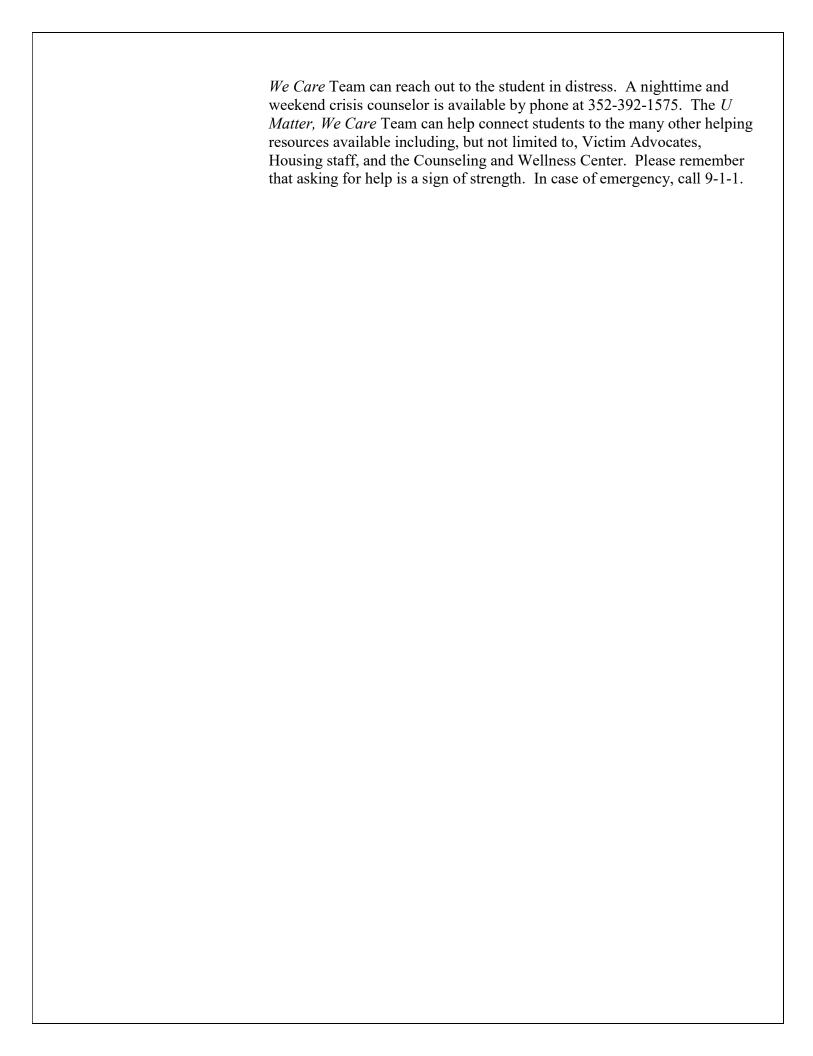
We, the members of the University of Florida community, pledge to hold ourselves and our peers to the highest standards of honesty and integrity. On all work submitted for credit by students at the University of Florida, the following pledge is either required or implied: "On my honor, I have neither given nor received unauthorized aid in doing this assignment." *All portions of the quizzes and lab reports are to be completed individually.*

Students with Disabilities

Students with disabilities requesting accommodations should first register with the Disability Resource Center (352-392-8565, www.dso.ufl.edu/drc/) by providing appropriate documentation. Once registered, students will receive an accommodation letter which must be presented to the instructor when requesting accommodations. Students with disabilities should follow this procedure as early as possible in the semester.

Additional Information

Your well-being is important to the University of Florida. The *U Matter*, *We Care* initiative is committed to creating a culture of care on our campus by encouraging members of our community to look out for one another and to reach out for help if a member of our community is in need. If you or a friend is in distress, please contact umatter@ufl.edu so that the *U Matter*,



List of Experiments

Laboratory 1	Introduction to CHM 4302L Laboratory Techniques
Laboratory 2	Cloning Strategy, Introduction to Restriction Enzymes, Agarose Gel Electrophoresis and the Polymerase Chain Reaction
Laboratory 3	Estimating the Concentration of the PCR Amplification Product by Gel Electrophoresis, PCR Clean-up, Digestion of the Expression Vector and PCR Product
Laboratory 4	Purification of DNA by Preparative Agarose Gel Electrophoresis and Purification of DNA from Agarose Gels
Laboratory 5	DNA Quantification of Purified hCA2 DNA and pETBlue-2 Vector DNA, Ligation of hCA2 DNA to pETBlue-2 Vector DNA and Transformation into <i>E. coli</i> NovaBlue
Laboratory 6	Screening pETBlue-2 Recombinants for hCA2 Insertion
Laboratory 8	Pilot Expression of Recombinant TunerTM(DE3) pLacI pETBlue-2 / hCA2
Laboratory 9	SDS-PAGE Analysis of the hCA2 Pilot-Scale Induction Experiment
Experiment 10	Expression and Partial Purification of Carbonic Anhydrase from Recombinant E. coli
Experiment 11	Gel Filtration Chromatography of Partially-Purified Carbonic Anhydrase
Experiment 12	Constructing a Purification Table for Carbonic Anhydrase Isolation and Kinetic Assays